Title: Bridging the gaps in test interpretation of SARS-CoV-2 through Bayesian network modelling **Authors:** Yue Wu¹, David Foley², Jessica Ramsay³, Owen Woodberry⁴, Steven Mascaro⁴, Ann E Nicholson⁴, Tom Snelling^{1,2,3,5,6}

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Extended abstract

Bayesian network (BN) models have been used within healthcare to bring clarity to complex problems.¹ In the absence of an established gold standard, an understanding of the testing cycle from individual exposure to test outcome report is required to guide the correct interpretation of SARS-CoV-2 (COVID-19) reverse transcriptase real-time polymerase chain reaction (RT-PCR) results and optimise the testing processes.² A wide variation in rates of false negatives has been reported, ranging from 1.8 to 58%;³ this variability may be attributable to heterogeneity in disease prevalence, patient age, timing of testing, type of specimen, other components of the pre-analytical phase and the RT-PCR assay employed across studies.⁴ We use the BN modelling approach to construct a comprehensive framework for understanding the real-world predictive value of individual RT-PCR results.

We elicited knowledge from domain experts to describe the test process through a facilitated group workshop. A preliminary model was derived based on the elicited knowledge, then subsequently refined, parameterised and validated with a second workshop and one-on-one discussions. Parameterisation of the model was conducted based on the literature and input from experts.

Causal relationships elicited describe the interactions of pre-testing, specimen collection and laboratory procedures, and RT-PCR platform factors, and their impact on the presence and quantity of virus thus the test result and its interpretation. As shown in Figure 1, the outcome model consists of 32 nodes in total, including 5 pre-testing factors (pink), 13 variables relevant to laboratory procedures (orange), 10 nodes (yellow) sequentially describing the trajectory of SARS-CoV-2 virus from the initial exposure event to final laboratory detection, and 4 summary nodes (blue) created for modelling purpose. 20 variables can be directly mapped to real-world observations therefore used as input variables if available, and the rest 12 variables are latent including the main output variable 'predicted classification' (node 31) which describes the probability of the classification being *truly positive*, *truly negative*, *falsely positive*, and *falsely negative*.

By setting the input variables as 'evidence' for a given subject, four scenarios were simulated to demonstrate potential uses of the model. In Figure 1, some input variables were entered to illustrate a scenario where a tested individual who has **negligible known recent viral exposure** (node **1**) and experiences **upper respiratory symptoms** (node **32**) but **tested negative** (node **30**), the probability of a *falsely negative* result is 14.6% (node **31**) if the person resides in a high viral prevalence setting (5%). The corresponding probability drops to 0.3% if it were a low viral prevalence setting (0.1%).

The core value of this model is a deep understanding of the total testing cycle, bridging the gap between a person's true infection status and their test outcome. This model can be adapted to different settings, testing modalities and pathogens, adding much needed nuance to the interpretations of results. This model requires validation with local, real-world datasets prior to application. We intend for future applications to integrate with other models that detail local epidemiological factors such as those developed by Fenton *et al.*⁵ to account for the complex and dynamic interactions between individual-level factors and population-level behaviours that influence the transmission and prevalence of SARS-CoV-2.

Figure 1. The testing model Intensity of most recent Days since most recent Age (11)
YoungChild 5.3% exposure (12) Trajectory of SARS-CoV-2 MoreThanSeven30.0% Heavy 0.0% 0.0% Light OlderAdult 28.3% LessThan24hrs 5.0% Negligible 100.0% Laboratory Days since first compatible symptom onset (13) 0.0% False 100.0% MoreThanSix20.6% ZeroToSix 79.4% NoSymptom 0.0% True 14.6% False 85.4% Body site sampled (15) Dyspnoea (33) Nasopharyngeal85.0% True 1.3% site (4) SalivaMouth LowerAirway 0.0% 5.2% Moderate 3.2% Faeces 10.4% Low Collection performance (17) Swab type (16) No 85.4% Flocked Good 91.7% NonFlocked 4.4% Poor 8.3% NotApplicable 9.9% Extraction process (21) Quantity of virus in sample (pre-transit) (5) utomated 99.0% Specimen quality (18) 1.0% 0.1% Good 91.8% Moderate 0.9% Poor 8.2% Operator performance (22) No 93.7% Good 95 0% Conditions of sample (post-transit) (6) Good 99.1% Extraction efficie 0.9% Poor Moderate 0.1% Match of primer to High99.7% Amplification process 0.9% Low target (25) (24)Low 0.3% 98.9% Low 99.7% Quantity of viral RNA in Poor 0.1% purified sample (7) High 0.1% Amplification efficiency (27) control (20) Moderate 0.1% Pass 100.0% Low 0.4% High 99.8% 99.4% 0.0% No Low 0.2% Shared target with Quantity of target in amplified sample (8) 0.1% 0.2% High threshold (29) control (28) Moderate 0.1% Pass 100.0% High 99.0% Low 0.2% 99.5% Detection of target (10) 0.0% Predicted classification (31) NotDetected 100.0% TrulyPos 0.0% report (30) TrulyNeg 85.4% ositive 0.0% alselyPos 0.0% 100.0% FalselyNeg 14.6%

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